



Evaluation of two high through-put (HTP) androgen receptor based assays: Utility of data for prioritization for further testing versus prediction of adverse effects.

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ABSTRACT

The androgen signaling pathway plays a critical role in sexual differentiation during development in mammals and is one of the better understood pathways in human development. Thus it was chosen as a model pathway to evaluate the potential of HTP in vitro assays as risk assessment tools. This study examined the interaction of chemicals with the androgen receptor (AR) using in vitro cell-based transcriptional activation assays. The chemicals identified as positive agonists or antagonists were then tested in competitive binding assays to confirm receptor interaction. An initial set of about sixty well-characterized compounds with varying affinities for the AR were tested. The in vitro results from these known compounds were compared to available data from in vivo Hershberger assays to evaluate the predictive capacity of the in vitro assays when compared to in vivo results. About fifty unknown chemicals were also tested in vitro and evaluated using the criteria developed. Results for chemicals with known activity in vivo and in vivo indicate that most with ED05s lower than 10⁻⁶ M were drugs or natural steroids. The pesticides and toxic substances known to have in vivo effects on the AR fell in the 10⁻⁴ to 10⁻⁶ M range. In the 10⁻⁴ to 10⁻⁶ M ED50 range, there was no correlation between in vitro activity and the in vivo potency. An examination of the chemicals in this range indicates that the limitations of the in vitro assays (failure to account for metabolic inactivation, activation, and half-life of a compound) result in a high rate of "false positives" precluding their use for accurate prediction of in vivo effects. However, since there were no "false negatives" (in vitro versus in vivo) these in vitro receptor assays can be used to prioritize chemicals for additional in vitro or short-term in vivo screenings for compounds that act via the AR signaling pathway in an HTP mode. Disclaimer: This abstract does not necessarily reflect EPA policy.

LIST OF CHEMICALS TESTED

Table with columns: Compound ID, Compound Name, CAS #, agonist, antagonist, binding, Compound #, Compound Name, CAS #. Lists various chemicals tested in the study.

OVERVIEW

Purpose: This extramural contract was designed to test the use of an in vitro pre-screening strategy as a method to prioritize chemicals for testing. Compounds were first tested in AR transcriptional activation (TA) assays for both agonist and antagonist activity. Positives were then tested in AR binding assays to confirm interaction with the receptor. Chemicals: Overall, about 125 chemicals were tested in three phases of work. Phase 1 - using 17 well-characterized compounds, the contractor established their proficiency with the assays. Phase 2 - Fifty compounds were tested. Most (46) were selected from the ICVAM list of Reference Substances for the Validation of In Vitro Assays plus four additional compounds of interest. Phase 3 - A total of 58 compounds from the ICVAM list were tested in either Phase 1 or 2. Phase 1 and 2 results were compared to in vivo results from the literature and used to set criteria for evaluation of Phase 3 unknowns. In Phase 3 - Fifty-seven unknown compounds were evaluated.

METHODS

Transcriptional Activation (TA) Assays: Androgen receptor mediated TA - MDA-Kb2. These cells have endogenous AR and stably express an androgen-responsive promoter (MTTV) linked to a luciferase reporter gene. Competitive Binding: AR Binding - Androgen Receptor-FP protocol (Androgen Receptor Competitor Assay (Invitrogen/Parvex)). Cytotoxicity: Cell viability in all cell based assays was monitored by propidium iodide (PI) uptake. Additional cytotoxicity assay (ATP assay) was performed for some Phase 3 compounds. Solubility: Limit of solubility was determined by a light scattering procedure using Nephelometry (Nepheloscan Ascent by LabSystems). Luciferase Interference: The two highest concentrations of each compound in Phase 3 were also tested for their ability to directly interfere with the luciferase enzyme itself (i.e. a non-receptor-mediated effect).

Phase 1 Notes: 1. Follow up work is needed. 2. Chemical is either a partial agonist or a mixed agonist and antagonist. 3. It is a false fit for antagonist (ED50) or binding (B).

Agonist example: Fluxymestrona 76-43-7. Antagonist example: Bicalutamide AR Antagonist. PHASE 3 PRIORITIZATION table with columns: Number, Compound Name, AR TA, AR binding, comments on cyt, priority, loc. CONCLUSIONS: The techniques used in this assessment can all be performed in an efficient high (or semi-high) through-put system. Having positive results of more than one in vitro assay adds confidence to the interpretation. Additional in vitro assay results (such as KI determination) may aid in further defining equivocal results. Results for chemicals from Phase 1 and 2 with known activity in vitro and in vivo indicate that most with ED50s lower than 10⁻⁶ M were drugs or natural steroids. An examination of the chemicals in this range indicates that the limitations of the in vitro assays (failure to account for metabolic inactivation, activation, and half-life of a compound) result in a high rate of "false positives" precluding their use for accurate prediction of in vivo effects. However, since there were no "false negatives" (in vitro versus in vivo) these in vitro receptor assays can be used to prioritize chemicals for additional in vitro or short-term in vivo screening for compounds that act via the AR signaling pathway in an HTP mode.